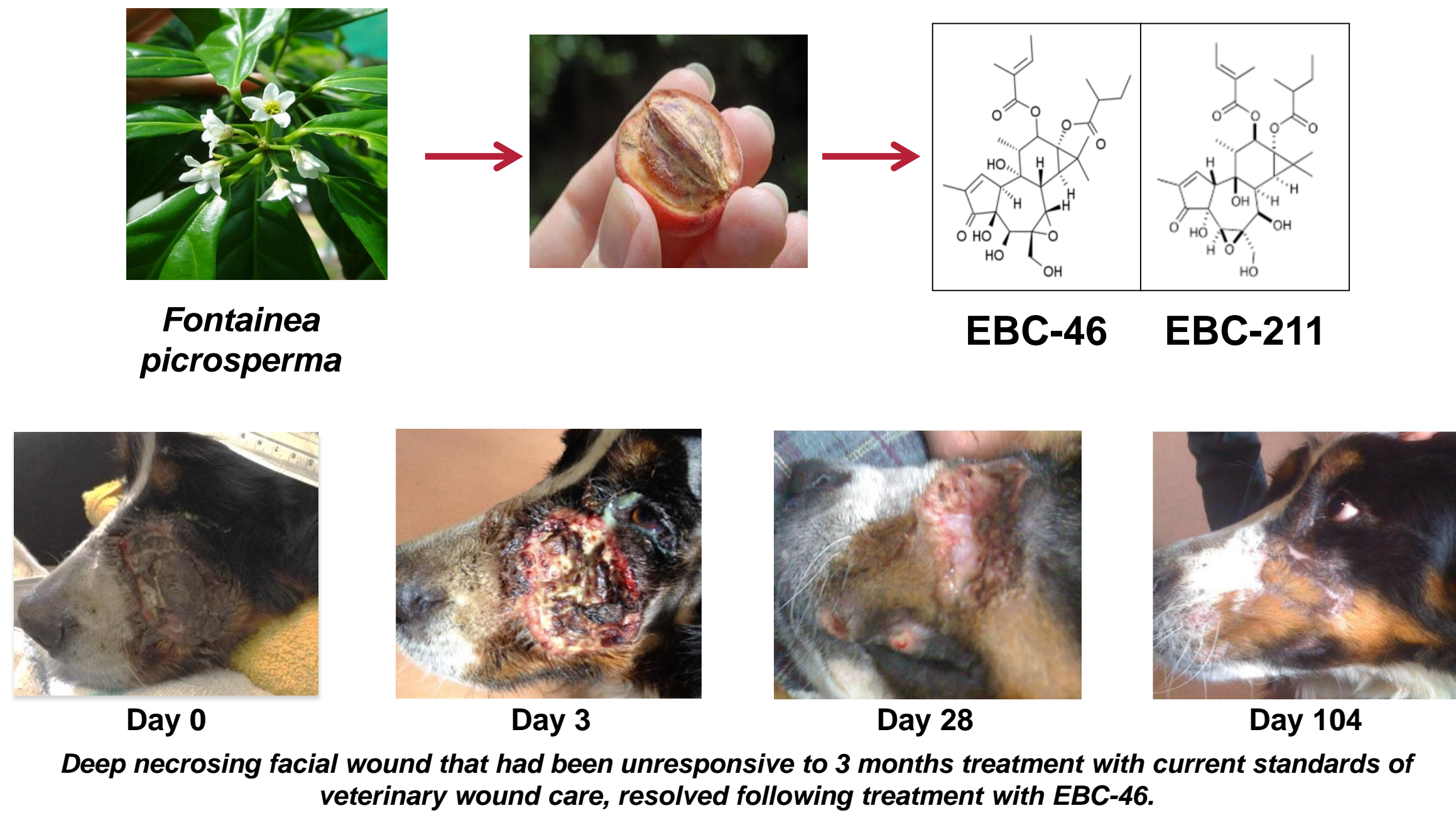


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Introduction

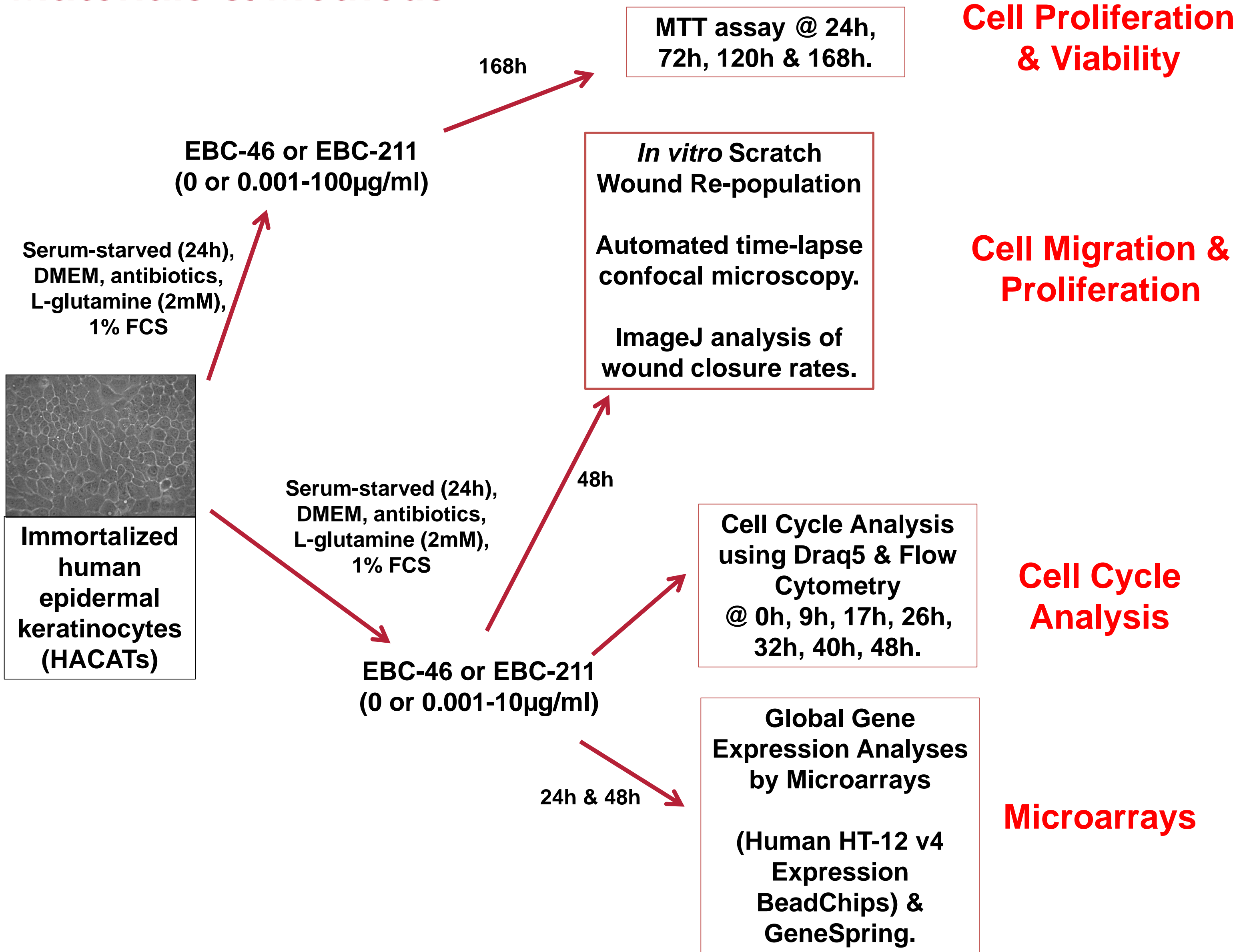
- The novel epoxy-tiglianes, 12-tigloyl-13-(2-methylbutanoyl)-6,7-epoxy-4,5,9,12,13,20-hexahydroxy-1-tigliane-3-one (EBC-46) & a less active related compound, 12-tigloyl-13-(2-methylbutanoyl)-5,6-epoxy-4,5,9,12,13,20-hexahydroxy-1-tigliane-3-one (EBC-211), occur within seeds of the Fontaine's Blushwood Tree, indigenous to Queensland's tropical rainforest¹.
- EBC-46 is currently being developed as an anti-cancer agent by Australian biotechnology company, QBiotics (www.qbiotics.com), for the intra-lesional treatment of cutaneous & sub-cutaneous tumours in humans & animals².
- In veterinary clinical trials, exceptional dermal wound healing responses, characterised by accelerated re-epithelialisation, closure & reduced scarring, have been consistently observed following tumour ablation by EBC-46².
- Such observations are reminiscent of the rapid re-epithelialisation rates evident during early gestational, foetal skin or oral mucosal healing³⁻⁴, leading to the potential of EBC-46 & EBC-211 being used to promote dermal re-epithelialisation during impaired healing chronic wounds or burn injuries.



Aims & Objectives

As little is known how epoxy-tiglianes promote wound re-epithelialisation in treated skin, we examined the effects of EBC-46 & the lesser active analogue (EBC-211), on keratinocyte proliferation & migration *in vitro* & the underlying mechanisms of action.

Materials & Methods



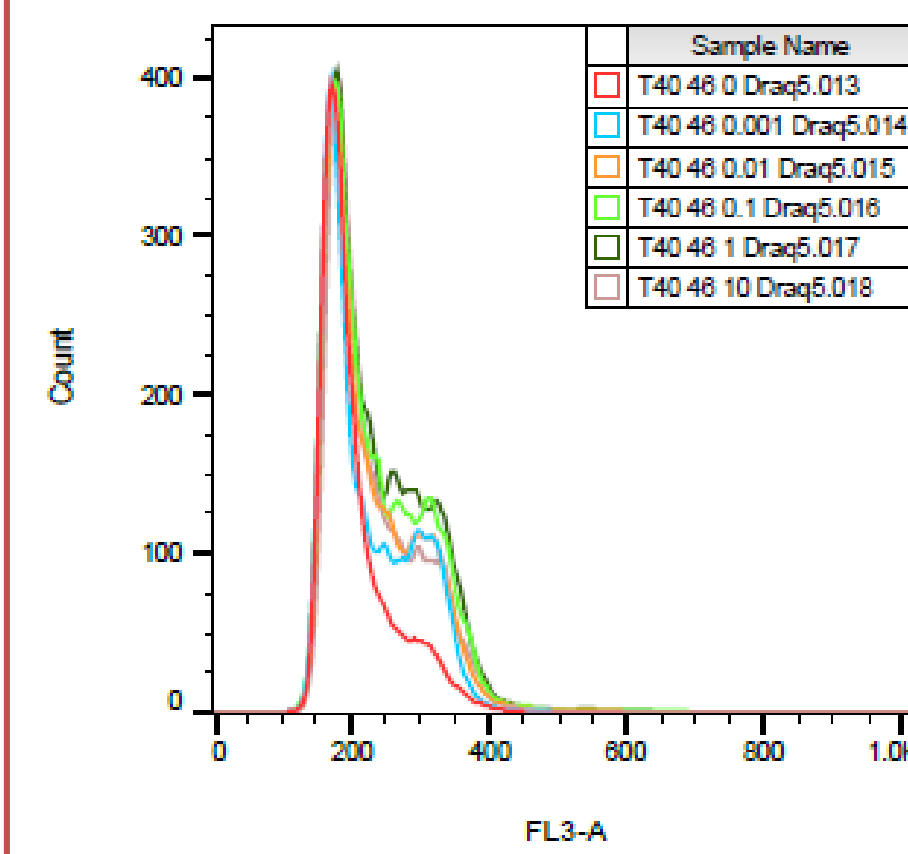
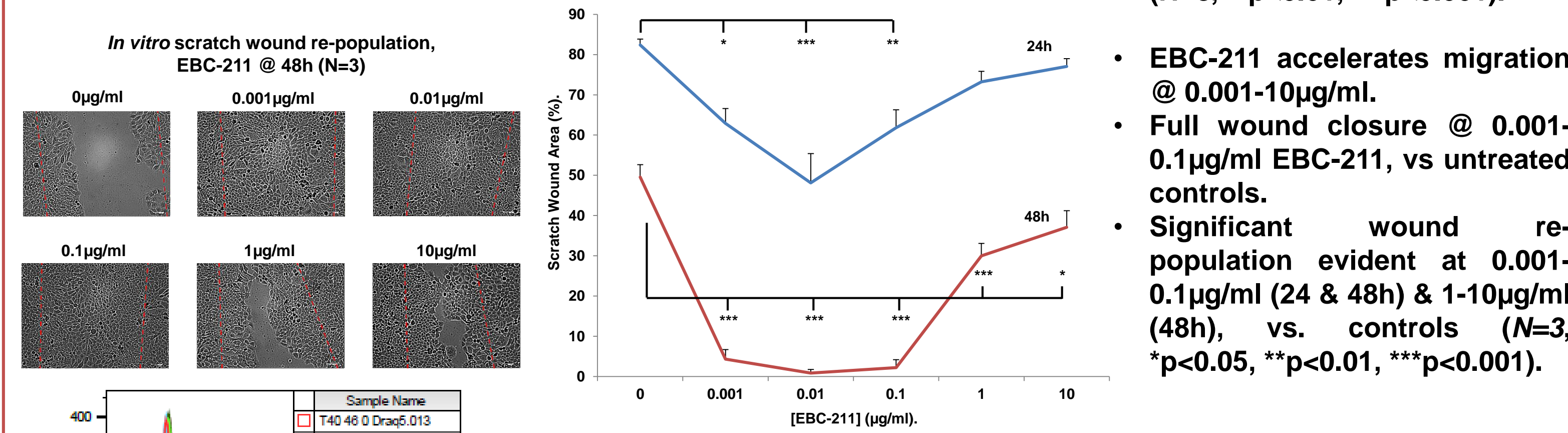
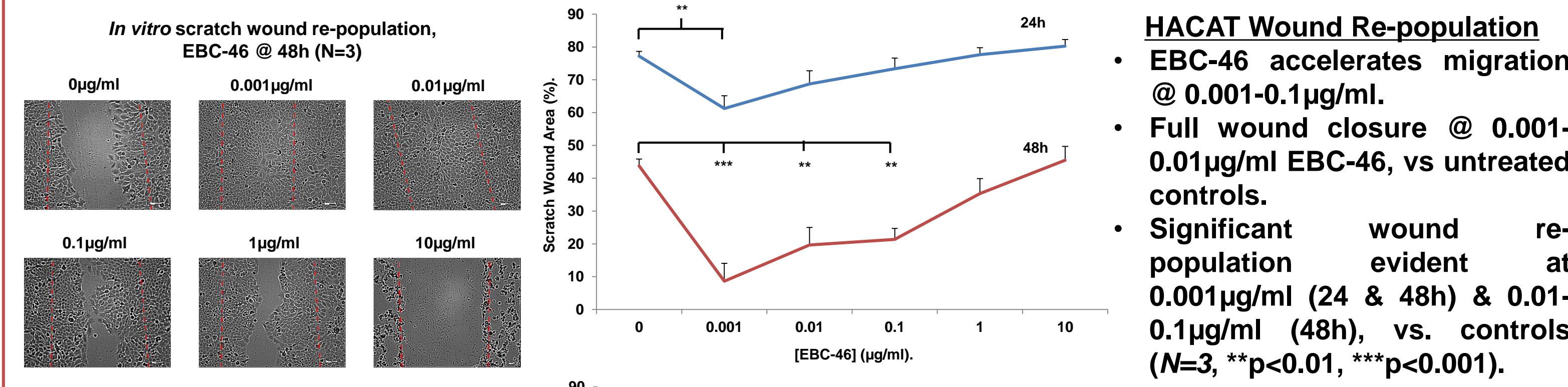
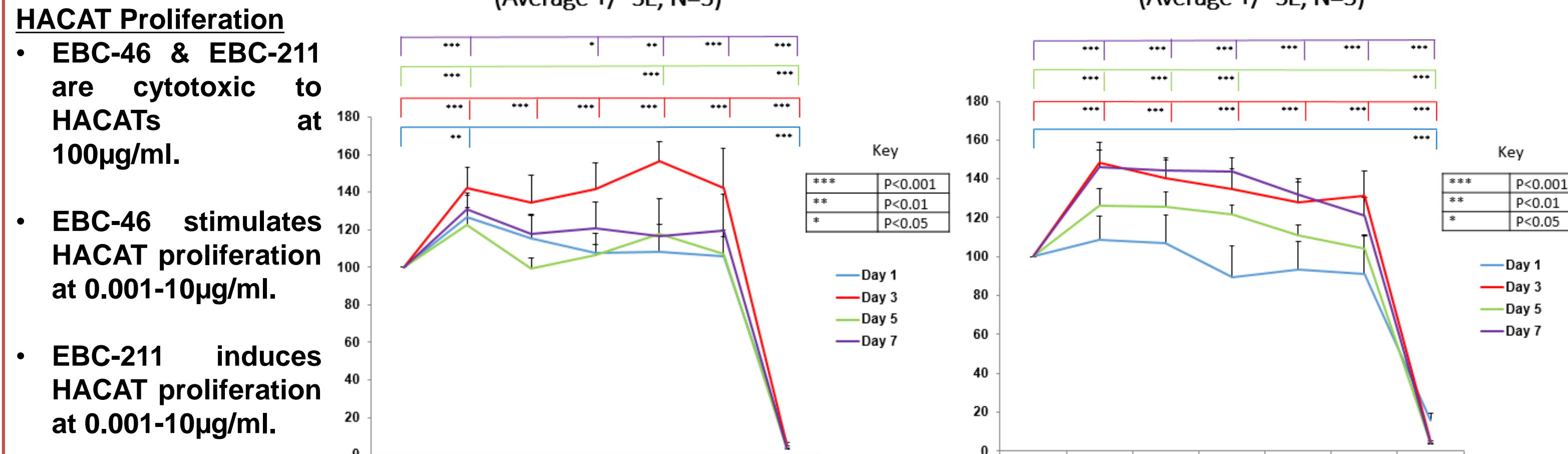
Conclusions

- Both EBC-46 & EBC-211 significantly increase HACAT proliferative & wound re-population responses *in vitro*.
- This study has also identified the genes principally involved in mediating enhanced keratinocyte proliferative & migratory responses following epoxy-tigliane treatment, which potentially occur by similar mechanisms to the favourable rapid healing events observed with epithelial wounds in the oral mucosa³⁻⁴.
- Such findings help explain the enhanced re-epithelialisation responses in epoxy-tigliane-treated skin & highlight their potential as novel therapeutics for impaired dermal wound healing situations.
- Further studies are elucidating the mechanisms by which epoxy-tiglianes induce these stimulatory effects.

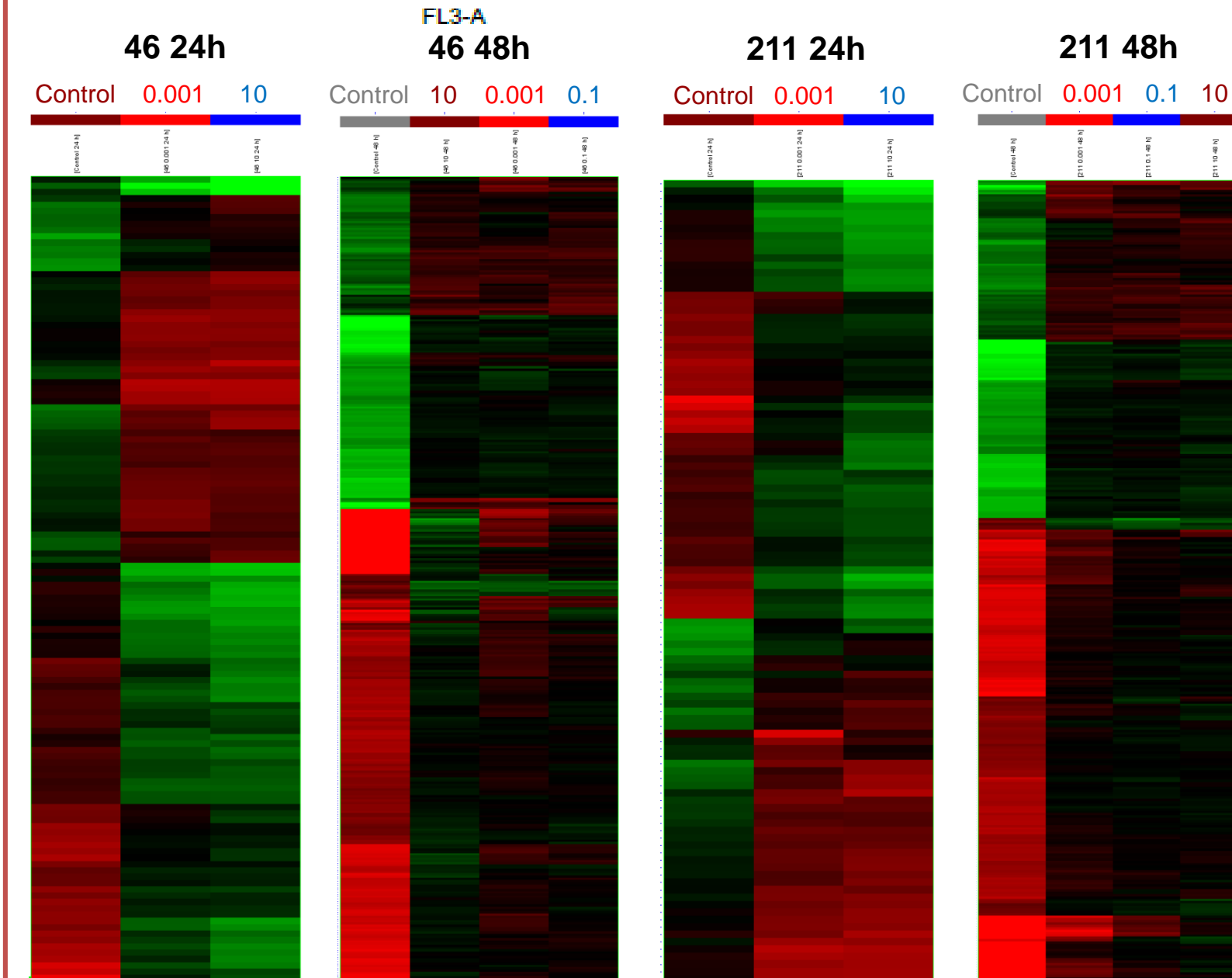
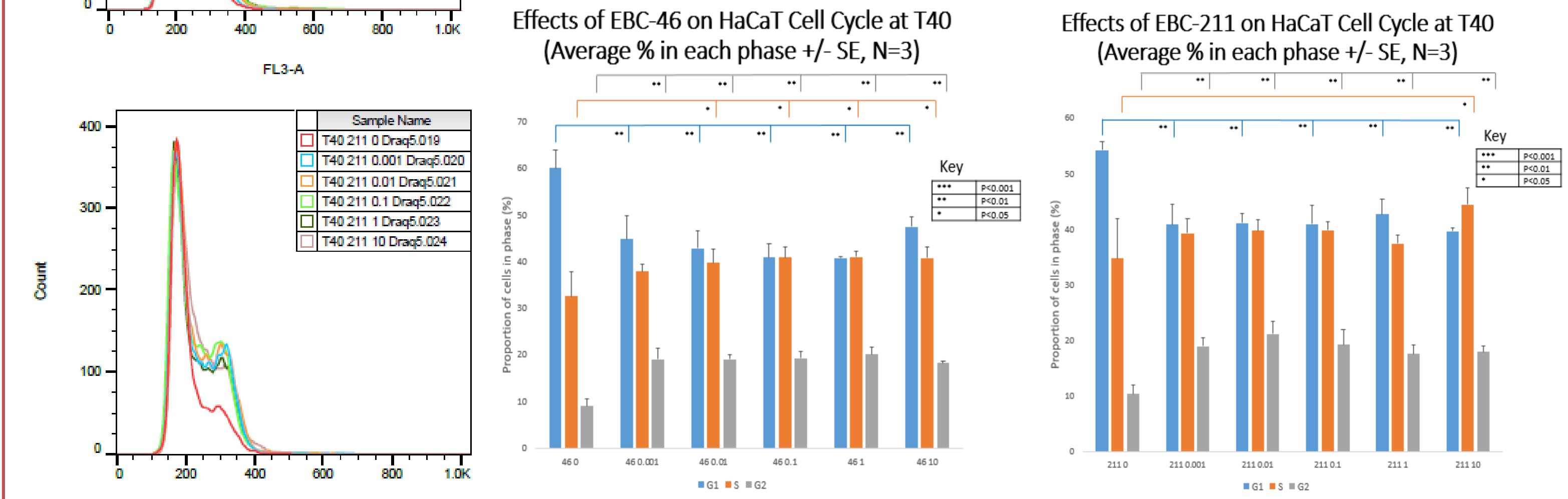
References

- Dong L *et al.* Anticancer agents from the Australian tropical rainforest: Spiroacetals EBC-23, 24, 25, 72, 73, 75 and 76. *Chem Eur J* (2009); 15:11307-18.
- Boyle GM *et al.* Intra-lesional injection of the novel PKC activator EBC-46 rapidly ablates tumours in mouse models. *PLoS One* (2014); 9:e108887.
- Yun-Shain L *et al.* Wound healing in development. *Birth Defect Res. C Embryo Today* (2012); 96:213-22.
- Glim JE *et al.* Detrimental dermal wound healing: What can we learn from the oral mucosa? *Wound Rep Regen* (2013); 21:648-60.

Results



- EBC-46 accelerates HACAT cell cycle progression @ 0.001-10µg/ml, compared to untreated control.
- EBC-211 accelerates HACAT cell cycle progression @ 0.001-10µg/ml, compared to untreated control.
- EBC-46 & EBC-211 treated cells progress out of G1 phase through S phase & into G2 phase ahead of the untreated control.



Fold change for untreated control against the five conditions for EBC-46 (A) & EBC-211 (B); 0.001µg/ml at 24h & 48h, 0.1µg/ml at 48h, & 10µg/ml at 24h & 48h.

Gene	Control vs EBC-211, 0.001 24h	Control vs EBC-211, 0.001 48h	Control vs EBC-211, 0.1 48h	Control vs EBC-211, 10 24h	Control vs EBC-211, 10 48h
KRT13	6.9	5.6	4.7		
KRT15	5.4	3.8	5.6		
KRT19	2.8	2.4	2.2		
KRT81	2.5				
KRT16			-3.2	-3.0	-4.0
KRT17		-2.3	-3.1	-3.0	-3.6
KRT6B			-2.89		-3.7
KRT17P3		-2.6	-3.3		-3.9
Cyclin B2	3.8	4.2	3.8		
Cyclin A2	2.9	3.0	2.7		
Cyclin B1	2.7	2.7	2.7		
CDKN3	3.2	3.4	3.1		
CDKN1A				-2.8	-2.1
MMP-1	4.1	6.0	4.7		6.2
GINS2	2.5	3.1	3.1	2.3	
POLE2	2.19		2.0	2.3	
UBE2C		3.6	3.6		3.7
PTHLH	2.7		2.6	2.9	2.3
IL-6		-4.3	-5.4		-6.2
IL-8		-2.8	-3.8	-3.2	-6.3
IL-32		-4.6	-6.9		-7.3
IFNB1	-3.3	-4.3	-5.1	-4.1	-4.4

- Heatmap visualization through hierarchical clustering of genes differentially expressed ≥ 2 fold by HACATs ($n=4$ biological repeats).
- Cultured in the presence of 0.001µg/ml, 0.1µg/ml or 10µg/ml EBC-46 (A) or EBC-211 (B); & compared to untreated controls at 24h & 48h.
- Data from the 4 biological repeats were grouped for combined analysis between treatments & vs the untreated controls.

Red = up-regulated genes, Green = down-regulated genes.

Gene	Control vs EBC-46, 0.001 24h	Control vs EBC-46, 0.001 48h	Control vs EBC-46, 0.1 48h	Control vs EBC-46, 10 24h	Control vs EBC-46, 10 48h
KRT13		5.3	5.3		6.3
KRT15		4.6	4.6	-2.8	5.7
KRT19		2.1	2.1		2.2
KRT81	2.3				
KRT16		-4.3	-4.3	-3.1	-4.1
KRT17		-4.1	-4.1	-3.1	-4.0
KRT6B		-3.2	-3.2		-3.5
KRT17P3		-4.3	-4.3	-2.5	-4.2
Cyclin B2		4.1	4.1		4.2
Cyclin A2		3.0	3.0		3.0
Cyclin B1		2.7	2.7		2.7
CDKN3		3.3	3.3		3.2
CDKN1A		-2.0	-2.0	-3.2	-2.2
MMP-1		5.8	5.8	2.5	5.1
GINS2	2.5	2.8	2.8	2.4	2.9
POLE2	2.4			2.41	
UBE2C		3.8	3.8		4.3
PTHLH	2.7	2.5	2.5	2.8	2.4
IL-6		-5.3	-5.3	-2.0	-7.2
IL-8		-5.6	-5.6	-4.4	-8.1
IL-32		-6.7	-6.7		-8.6
IFNB1	-3.8	-4.3	-4.3	-4.2	-5.0

- Up-regulated genes included certain keratins (KRT9, KRT13, KRT15, KRT81), positive cell cycle & proliferation regulatory genes (CCNB2, CDKN3, CDCA7, GINS2, KIAA0101); & proteinases (MMP-1, MMP-7, MMP-10).
- Down-regulated genes included other keratin (KRT6B, KRT16, KRT17) & cytokines (e.g. IL-6, IL-8, IL-32).

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This work is covered by filed patent, WO2014169356.

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